NIH Institutional Biosafety Committee Minutes
Location: National Cancer Institute at Frederick Facilities

August 6, 2026
12:00 PM – 2:00 PM
Virtual Meeting available via WebEx

Members Present:

Voting Members (Quorum = 8)	Voting Members (continued)	*Ex officio* (non-voting)
☒ Sharon Altmann (Community member)	☒ Stephen Hughes	☐ Samuel Denny
☒ Theresa Bell	☒ Dan McVicar (Chair)	☐ Krisztina Janosko
☒ Stella Spears (Alternate for J. Gulani)	☐ Bradley St. Croix	☐ Michael Kujawa
☒ Eric Freed	☒ Brandon Keele	Other/Guests (non-voting)
☐ Jatinder Gulani	☒ Valentina Perepnikhatka	☒ Ligia Pinto
☒ Effie Nomicos (Community member)	IBC Office (non-voting)	☒ Troy Kemp
☒ Sarah Hooper	☒ Wendy Kitta	
☒ Jason Yovandich		

Announcements & Call to Order

I. Meeting called to order by IBC Chair at 12:05 pm.

Review of Past IBC Meeting Minutes

I. June 11, 2026, Minutes
   a. Comments on minutes: None provided.
   b. The minutes were unanimously approved as written.

New Committee Business

I. BSO or IBC lead reviewer preliminary registration approvals since the previous meeting:

   a. Pathogen only Registration Numbers: Not applicable.

   b. rDNA Registration Numbers:
      - Dr. Shari Price- IBC 2026-38 (Ref# 200515): ITL-Investigative Toxicology Laboratory (New, Storage-Only)
      - Dr. Claudia Palena- IBC 2026-39 (Ref# 200251): Breeding of C57Bl/6 BALB/c and B6-CEA Transgenic mice (Renewal, Breeding-Only)

c. Registration amendments (non-administrative)
   - ❖ Dr. Leslie Aldrich: 2024-41-A6. *Description:* Add plasmids for the generation of viral vectors that will be used for human cell line transduction and updates to the staff roster.
   - ❖ Dr. Alex Compton: 2024-28-A1. Description: Remove projects involving viral work, which are no longer pursued.
   - ❖ Dr. Robin Dewar: 2024-23-A2. Description: Add a new neutralization assay for diagnostics.
   - ❖ Dr. Vinay Vyas: 2025-17-A1. Description: Add a new plasmid and updates to the staff roster.

d. Additional Committee Discussion: N/A

II. Registrations for Committee review

Ref# 200077 (Renewal), Dr. Colin Wu: Translational Control of Gene Expression

i. *Reviewers:* Altmann, Hughes

ii. *Summary:* The laboratory investigates the molecular mechanisms of ribosome quality control, focusing on how cells detect defective ribosomes, aberrant translation events, and translational stress. Approximately 10– 15% of human cancers harbor chromosomal deletions at specific loci, which frequently result in the co-deletion of a gene that encodes for a critical component of RNA and ribosome surveillance pathway. A major goal of the research is to explore the unique vulnerabilities of gene-loss cancers, identify actionable therapeutic targets, and develop therapeutic strategies for cancers with defects in surveillance pathways.

iii. Committee Discussion:

-Section 4.2.1.1: Regarding the attached file: Please clarify line items 13, 14, 24, and 25 to reflect that the listed vectors carry either guide RNAs or shRNAs and that none of the vectors carry the coding information for the oncogenes or tumor suppressors.

-General comment about the attached file "CW cell culture and viruses SOPv5": Please modify the text on page 4, bullet point 9.2d. to eliminate the statements that do not apply to the research undertaken under the scope of this registration, namely, the work with dangerous inserts within the viral vectors, as discussed during the meeting with the IBC Chair. Please also make sure to include cautionary statements for staff, to alert them about potential hazards pertaining to the viral vector work.

iv. Animal studies proposed: No

v. Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.

vi. *Work is approved at:* BSL1, BSL2

vii. Training prescribed to staff in the roster: BBP, VVST

viii. *Relevant sections of the NIH Guidelines:* Section III-D-1-a.

ix. Public comment: None provided by the general public.

x. Votes: 11 approved, 0 abstained

xi. The committee unanimously voted to defer review to the assigned lead reviewers. If it is determined that oncogenes, proto-oncogenes or tumor suppressors are worked with, the registration must undergo full committee review once more.

Ref# 200052 (Renewal), Dr. Daniel McVicar: Retroviral and Adenoviral Manipulation of Metabolism and Inflammation

i. *Reviewers:* Altmann, Hughes

ii. *Summary:* Lentivirus-mediated delivery of shRNA provides a robust method for stable gene knockdown, allowing sustained suppression of target gene expression in dividing and non-dividing cells. These studies enable functional characterization of genes involved in tumor initiation, progression, metastasis, and drug resistance. By identifying genes that drive malignant phenotypes, this approach supports the discovery and validation of molecular targets for the development of more effective cancer therapeutics.

iii. *Committee Discussion:*
-Section 4.2.1.1: Under column "Gene Type Encoded within Vector": Please indicate the viral envelope being used for each applicable entry.

-Section 5.34 and 7.13: Please modify the text to indicate that paper towels are first placed over the spill and the freshly prepared disinfectant solution is then gently poured over the paper towels (poured from the outer perimeter, towards the center of the spill).

-Section 7- General Comment: Please describe the murine cells that are obtained from the lavage procedure. Details about these primary cells should be described throughout section 7 along with the mouse bone marrow cells.

-Section 7.10: Please update the text to also include mouse-derived materials.

iv. *Animal studies proposed:* Yes, mice.

v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is approved at:* BSL2, ABSL1, ABSL2

vii. Training prescribed to staff in the roster: BBP, AEP, VVST

viii. *Relevant sections of the NIH Guidelines:* Section III-D-1-a

ix. Public comment: None provided by the general public.

x. Votes: 10 approved, 1 abstained.

xi. The committee voted to approve, pending minor clarifications. Dr. McVicar recused.

Ref# 200057 (Renewal), Dr. Lisa Ridnour: Role of Nitric Oxide in Breast Cancer Disease Progression

i. *Reviewers:* Gulani, St. Croix

ii. *Summary:* The laboratory has devoted the last decade examining the roles of genes involved in tumor expression during immunosuppression and the disease progression of aggressive forms of breast cancer including Estrogen receptor negative (ER-) and triple negative breast cancer (TNBC). Collectively, these studies show that dual treatment with targeted inhibitors significantly enhances adaptive immunity and immune memory with and without therapeutic treatment. Therefore, the targeted inhibitors could offer clinically available alternatives for patients who have otherwise exhausted all treatment options. This work is supported by the results of past and ongoing clinical trials where patients are treated with similar inhibitors and other FDA approved drugs.

iii. Committee Discussion:

-Section 4.2.1.1:

1. Regarding the last two line items listed on the table: It isn't clear whether the last 2 lentiviral vectors carry CRISPR (or exactly what they carry, for example, do they carry oncogenes, proto oncogenes, tumor suppressor genes, etc.). Please clarify. If CRISPR is carried, please enter a statement under column "Gene Type Encoded within Vector" indicating that CRISPR is genotoxic.

2. Under column "Transduced cells administered to Animals" for the last three line items: Please clarify whether the cells have remained in cell culture for at least 10 days, prior to them being injected into animals.

3. Under column "Vector Used for Transduction? If yes, identify cells": The entries currently indicate that MB231 and 4T1/4T1 CRISPR are transduced. However, section 6.19 of the form indicates that 4T1/4T1 CRISPR and EO771 are transduced with viral vectors. Please correct as applicable to ensure that both sections are consistent.

-Section 5.5:

1. Please clarify the first sentence of the text to indicate that all work with human cell lines and tissues is always conducted in a BSC while following BSL2 practices (not only in situations where there is potential for aerosol generation).

2. Please indicate in the text whether the sonicator is located inside a BSC or other primary containment device. Please make sure to identify the engineering control being used to contain the sonicator.

3. Please attach an applicable SOP that describes the rules of the laboratory and how the work with the cell lines and tissues is conducted safely.

Section 5.19.1.1:

1. Please correct the text to replace transfection/transfected with transduction/transduced.

2. Please specify in the text, for how long the cells remain in culture post-transduction, before being injected into animals. Do the cells remain in culture for at least 10 days post-transduction?

-Section 5.20.1.1: Please include a disclaimer in the text, indicating that human tissue recovered from mice may be infected with, and produce, murine retroviruses that will infect and replicate in human cells.

-Section 9.5: Are the BSC and incubator (where the cells requiring the use of the toxin material, are manipulated and grown) identified with signage to indicate that toxin material is present? Please address in the text.

-Section 9.8: Please indicate in the text whether an inventory of the toxin material is maintained and kept up to date.

iv. *Animal studies proposed:* Yes, mice.
v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is described at:* BSL2, ABSL1, ABSL2
vii. Training prescribed to staff in the roster: BBP, AEP
viii. *Relevant sections of the NIH Guidelines:* Section III-D-1-a.
ix. Public comment: None provided by the general public.
x. Votes: 10 approved, 1 abstained.
xi. The committee unanimously voted to conditionally approve pending clarifications. Dr. McVicar recused.

Ref# 200087 (Renewal), Dr. Christine Heske: Use of Pediatric Solid Tumor Cell Lines and PDXs for in vivo Studies

i. *Reviewers:* Altmann, Gulani
ii. *Summary:* Rhabdomyosarcoma (RMS) is the most common soft tissue sarcoma in children, and it requires novel treatment approaches. Currently, the mainstays of treatment are toxic chemotherapy agents, radiation, and surgery and despite this, a subset of patients who have high-risk disease do not survive. Further, for those who do survive, these treatments result in long-term toxicities. Thus, identification of less toxic agents that can target tumor cells while potentially sparing normal cells are necessary. The objectives of this study are to evaluate promising agents identified through work in cancer cell lines and in animal models of this disease, in preparation for potential translation to the clinic.

iii. Committee Discussion:
-Section 5.7: Please enter a statement acknowledging that human cells implanted in mice may come out harboring viruses that can infect humans.

iv. *Animal studies proposed:* Yes, mice.
v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is described at:* BSL2, ABSL2
vii. Training prescribed to staff in the roster: BBP, AEP
viii. *Relevant sections of the NIH Guidelines:* N/A- No recombinant material identified.
ix. Public comment: None provided by the general public.
x. Votes: 11 approved, 0 abstained.
xi. The committee unanimously voted to approve pending minor clarification.

Ref# 200097 (Renewal), Dr. Terrence Yamaguchi: The Role of Wnts in Vertebrate Development and Cancer Molecular Clones

i. *Reviewers:* Hughes, St. Croix
ii. *Summary:* The laboratory studies specific gene signaling during embryonic development in mice. The laboratory utilizes mouse knock outs or conditional deletion of genes to understand how abnormal gene function leads to disease states. The over-expression of specific genes is also studied to look at downstream targets. Mouse embryonic stem cells (ESCs) are used in-vitro to model developmental

events in mice. Additionally, the laboratory studies whether the developmental mechanisms elucidated in mouse ESCs and mouse models can apply to human cells.

iii. Committee Discussion:

-Section 4.2.1.1: The text regarding the gRNAs is confusing as gRNAs do not function as oncogenes or tumor suppressors. Please clarify the language.

-Section 6.12: There are two primary reasons VSV-G cannot replace env in a replication competent retrovirus. 1) the env gene also encodes, in alternate reading frame, other proteins HIV needs. 2) high levels of VSV-G are toxic to cells. Cells that make lots of VSV-G die. That appears to be fine for VSV, but it isn't OK for a retrovirus. Please update the text to reflect these points.

-Section 7.12: Section 7.9 only identifies one BSL2 facility; however, the text in this section references a BSL1 facility as well. Please correct to ensure that both sections are consistent.

iv. *Animal studies proposed:* Yes, mice

v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is described at:* BSL2

vii. Training prescribed to staff in the roster: BBP, AEP

viii. *Relevant sections of the NIH Guidelines: Section III-D-2-a.*

ix. Public comment: None provided by the general public.

x. Votes: 11 approved, 0 abstained.

xi. The committee unanimously voted to approve, pending clarifications.

Ref# 200141 (Renewal), Dr. Parirokh Awasthi: Production of Genetically Engineered Mice

i. *Reviewers:* Bell, Perepnikhatka

ii. *Summary:* The Laboratory only accepts projects that are strictly classified under Biosafety Level 1 (BSL1), Biosafety Level 2 (BSL2) and Animal Biosafety Level 1 (ABSL1) defined within the CDC/NIH publication (Biosafety in Microbiological and Biomedical Laboratories; 6th edition). The objective is to generate genetically engineered mice for the NIH scientific community

iii. Committee Discussion:

- Section 6.10: Please correct the response to this question to state “Yes”.

iv. *Animal studies proposed:* Yes, mice

v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is approved at:* BSL2, ABSL1

vii. Training prescribed to staff in the roster: BBP, VVST

viii. *Relevant sections of the NIH Guidelines:* Section III-E-3

ix. Public comment: None provided by the general public.

x. Votes: 11 approved, 0 abstained.

xi. The committee unanimously voted to approve pending minor clarification.

Ref# 200241 (New), Dr. Ligia Pinto: Determination of Biomarker and Antibody Levels within Human Specimens Using Luminex- based Technology.

i. *Reviewers:* Altmann, Perepnikhatka

ii. *Summary:* The objective of this protocol is to test human specimens using Luminex bead-based technology to determine cytokines and antibody levels and evaluate the associations of the cytokines and antibody levels with age, gender, vaccination status, natural infection status, and other comorbidities. Testing will be performed according to manufacturer's procedure manual or internal developed standard operating procedures (SOP)s for each Luminex bead based assay.

iii. Committee Discussion:

-Section 7.5: The discussion during the August meeting focused on the processing of clinical specimens, which occurs in the BSL2 laboratory setting. The NCI at Frederick IBC is required to uphold the OSHA Bloodborne Pathogens Standard, which provides the basis for the oversight of the work with clinical specimens.

During the discussion, the PI indicated that HPV is not known to circulate in the bloodstream, and that it is localized in epithelial cells within the cervical region, thus presenting a lower risk. For this reason, the PI proposed the use of additional PPE for situations where the HPV samples cannot be entirely processed in the BSC. The OSHA regulation does not specifically require the use of a BSC for standard practice in a clinical setting. However, the IBC requires enhanced PPE to be incorporated into current laboratory practices to protect staff. Currently, laboratory staff don safety glasses, gloves, lab coats.

In addition to the standard PPE described above, the IBC requires the use of a face shield when transporting covered plates (which are subsequently uncovered at the bench) from the BSC to the bench for the washing step on the microplate washing equipment.

Upon completion of the washing step, the plate needs to be covered and returned to the BSC for further processing. The face shield must be worn continuously, during the transport of plates to the BSC as well. If multiple washing cycles are conducted, the face shield must not be removed to avoid cross contamination resulting from the handling of the face shield with contaminated gloves. Please update the text in the registration and in the associated SOPs to reflect these requirements.

iv. *Animal studies proposed:* No

v. Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.

vi. *Work is described at:* BSL2, BSL2*

vii. Training prescribed to staff in the roster: BBP

viii. *Relevant sections of the NIH Guidelines:* N/A-No recombinant material identified.

ix. Public comment: None provided by the general public.

x. Votes: 11 approved, 0 abstained.

xi. The committee unanimously voted to approve, with the provision that plates are removed to be unsealed and washed outside the BSC and then returned to the BSC for processing, while staff continuously wear a face shield (in addition to standard PPE).

Ref# 200317 (Renewal), Dr. Jairaj Acharya: Sphingolipid and Phospholipid Signaling Section

i. *Reviewers:* Hughes, Yovandich

ii. *Summary:* Sphingolipids drive fundamental processes in biology ranging from cell division and differentiation to apoptosis. In order to understand the biological significance of individual enzymes/proteins in sphingolipid biology, the laboratory has been engaged in the last two and half decades in addressing their role in vivo using Drosophila and mouse as model organisms and complementing these efforts using ex vivo and biochemical studies. To this end, the laboratory will be generating mutations in the genes of sphingolipid metabolic pathway in vivo in the mouse and Drosophila. Additionally, the laboratory will conduct knockdown and overexpression studies using tissue culture cells including human tissue culture cell lines. The objective of the study is to generate DNA vectors that will aid in the identification of interacting partners of various sphingolipid biosynthetic enzymes.

iii. Committee Discussion:

- Section 3.2.1.1:

1.Under column "Gene Type Encoded within Vector", regarding the pRetroX entry:

a. Recombination is a potential problem if RetroX is used in murine cells. Please acknowledge this in the text.

b. Please confirm the viral envelope that is being used with the vector.

2. Please generate and attach a consolidated Excel spreadsheet that shows the human cell lines and animal cell lines that are transduced with viral vectors. The committee is seeking a comprehensive list that clearly conveys which cell line/viral vector/cargo are introduced in vivo vs. only used in vitro.

-Section 4.18.1.1: There is a possibility that RD114 env could, if introduced into MLV, produce a replication competent virus that would infect human cells. A definitive answer to this question could not be found in PubMed at the time of the review. In the absence of data, this combination should be treated with extra caution, and the potential hazard needs to be acknowledged in the text.

-Section 5.2: Details about primary cells should be discussed in section 6, in lieu of section 5. Please remove all references to them from section 5 and enter all the relevant information in section 6.

-Section 5.3: Please select "Not Applicable" as the response to this section.

-Section 5.5:

1. Regarding the second paragraph: The text in the first sentence of the text describes the laboratory's work in gene expression and knockdown studies utilizing viral and non-viral vector systems in animal cells. The second paragraph then proceeds to describe cultured lines being injected into animals. These statements could be construed as if transduced murine cells are administered in vivo. This is in direct conflict with the viral vector summary table in section 3 of the form, and with the various lists of viral vectors attached to the registration (which explicitly state that transduced cells are not administered in vivo). Please correct the text in this section to ensure consistency with the viral vector lists. Note: Section 5.7 also states that no in vivo work will occur.

2. Please remove the last sentence of the text from this section, as the work with primary cells only needs to be described in section 6 of the form.

3. The attached Virus SOP also describes the possibility for injections into mice (which also conflicts with statements made in other sections and in the viral vector lists). If this no longer applies to the work scope described in this registration, please remove those references from the text.

-Section 5.12: Regarding the statement "There is a possibility for recombination events to occur, namely, the recombination of endogenous elements within animal cells/animal tissues and the viral vectors...": Please remove the reference about animal tissues. Other sections (and attachments) indicate that no transduced cells are injected in vivo.

-Section 5.13:
1. In terms of the last sentence, the MLV vectors can be mobilized recombine at any time. 10 days would allow the initial virus stock to infect or be inactivated. Please clarify the text accordingly.

2. In the last sentence, please remove the statement "before injection" (as no in vivo work is identified with transduced cells). Please replace that statement with "before in vitro procedures", for clarity.

-Section 5.16.1.1: The file "Safety protocol for lentiviral and adenoviral work in the Acharya laboratory" references the possibility for in vivo work. Please update the file to remove the references.

-Section 5.31: Please remove the second portion of the text, which relates to animal work and tissue dissection. Previous sections of the form indicate that no in vivo work is performed with the cells described in section 5.

Section 5.32:
1. Please correct the text to indicate that ethanol cannot be used on its own as a disinfectant. Either Cavicide or freshly prepared bleach solution need to be used for the applications described in section 5.

2. Please delete the last paragraph.

-Section 5.33: Please remove all references about tissues.

-Section 5.35: In the last paragraph: Remove all references to animal facilities.

-Section 5.36: Please delete the second paragraph of the text (only leave the first sentence as the response).

-Section 5.37: Please remove the text from this section, and indicate "N/A" as the response.

-Section 6.5:
1. Please expand the text to provide a comprehensive description about how the bone marrow cells are modified with viral vectors prior to being introduced into animals. Additionally, a safety-focused laboratory SOP that describes these procedures needs to be attached to this section and referenced in the text.

2. The statement following statement needs to be clarified further: "We will Identify genes that interact with sphingolipid metabolizing enzymes using shRNAs that will be introduced into mouse fibroblasts or T cells or hematopoietic stem and progenitor cells using replication defective lentiviral vectors or retroviral or adenoviral vectors. Occasionally, we intend to use Adeno-Cre and viral vectors and lentiviral vectors to modify established MEFs and ES cells". Please make sure to address the following:
- Which specific vectors are used with which primary cell types?
- Which of those cells are transduced and subsequently introduced in vivo?
- Which cells are specifically modified with RetroX vectors? Are those cells injected in vivo? If yes, there needs to be a statement highlighting the hazards associated with the vector (in the context of dangerous cargos and the potential for recombination).

-Section 6.6.1.1:
1. Please clarify where the Isolation of Bone marrow, the preparation, cell transduction and transplant will occur. Please identify whether the procedures will take place in the PI's lab versus the animal facility. Also, are all steps performed in the BSC?

2. Where will the harvest of cells/tissues and blood collection take place (PI's laboratory vs. animal facility)? In the BSC?

3. Will animals be anesthetized for all injection procedures? Will all injections be performed in the BSC?

4. Under "Preparation of cells to be transplanted": Please clarify for how long in total the transduced cells remain in culture prior to injection in vivo. Do they remain in culture for 10 days or more in culture prior to injection?

-Section 6.6.1.3: Please complete this table.

-Section 6.11: Please clarify the text to indicate that animal carcasses are not autoclaved. They are to be bagged and placed in a facility dedicated freezer for eventual pick up by EHS Waste Management.

-Section 6.15: The text about needle disposal is slightly ambiguous. Please clarify that needles are disposed of in sharps container located inside the BSC, not outside.

-Section 7.3: Please specify if the bacteria are heat-killed.

-Section 7.9: Please complete this table.

- Section 7.12: Is the bacterial strain not expected to cause disease in susceptible employees (such as immunocompromised status) because the bacteria have been previously heat killed or inactivated? Please provide an explanation in this section. The response cannot be "N/A".

-Section 7.16: Please respond to this section and complete the follow up text boxes that may be generated.

-Sections 7.19 through 7.20: Please provide the required information in these sections. The responses cannot be "N/A".

-Section 8.8:
1. Please describe in the text how access to the storage location is restricted.
2. Please indicate in the text whether an inventory is maintained and kept up to date.

-Section 8.16:
Please enhance the text by providing a description about evacuation procedures for situations where spills happen outside of primary containment. The area needs to be evacuated for 30 minutes to allow aerosols to settle down. Signage is then posted on the door to inform other employees to avoid entry into the space. After the 30 minutes waiting period has elapsed, employees don appropriate PPE and initiate spill clean-up activities.

-Section 9.11: Please complete all fields on this table.

-Section 9.15: Previous sections only mention microinjections but not actual work with grown mice. If that is correct all references to the animal facility should be removed.

iv. *Animal studies proposed:* Yes, mice.
v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is described at:* BSL1, BSL2
vii. Training prescribed to staff in the roster: BBP, AEP, VVST
viii. *Relevant sections of the NIH Guidelines:* Section III-D-4, Section III-E-1, Section III-E-3
ix. Public comment: None provided by the general public.
x. Votes: 11 approved, 0 abstained.
xi. The committee unanimously voted to defer review to the next available IBC meeting.

Ref# 200518 (New), Dr. Ligia Pinto: Long Non-Coding RNA (lncRNA)

i. *Reviewers:* Freed, Perepnikhatka
ii. *Summary:* The objective of this research is to evaluate the biological activity and therapeutic potential of a gene encoding a short fragment derived from a liver-expressed long non-coding RNA, which is associated with lipid metabolism and gluconeogenesis. This work is relevant to metabolic diseases involving abnormal glucose and lipid regulation.

iii. Committee Discussion:
This registration is well described and has undergone previous reviews that led to the organization of the attachments and administrative clarifications in the language.

iv. *Animal studies proposed:* No.
v. *Committee discussion about Dual-Use Research of Concern (DURC) and enhanced Potential Pandemic Pathogen (ePPP) considerations: There were no DURC or ePPP concerns with the described experiments.*

vi. *Work is described at:* BSL2
vii. Training prescribed to staff in the roster: BBP
viii. *Relevant sections of the NIH Guidelines:* Section III-D-1-a.
ix. Public comment: None provided by the general public.
x. Votes: 11 approved, 0 abstained.
xi. The committee unanimously voted to approve this registration.

III. Registration amendments for full committee review:
i. None
ii. Reviewers: N/A
iii. Review Summary: N/A
iv. Committee Discussion: N/A
v. Animal studies proposed: N/A
vi. *Committee discussion about the dual-use and ePPP potential of these experiments:* N/A
vii. Work is approved at Biosafety Level(s): N/A
viii. Relevant sections of the NIH Guidelines: N/A
ix. Committee vote/recommendations: N/A

IV. Committee Review of Inactivation Procedures (if not reviewed under a registration)
a. None

V. Standard Operating Procedures/Plans
a. None

VI. Serious Adverse Events in Clinical Trials reviewed by the Committee
a. Not applicable

VII. Committee Training
a. None

Reports

I. Biosafety Officer Report –
a. *Report I:* The Biosafety Officer presented a near miss involving a spill in a biological safety cabinet. Blood was spilled in a laboratory; however, all materials were contained in the biosafety cabinet and no staff exposure occurred.

II. Other reports
a. None

Around the Room/Committee Discussion

I. None

Adjournment

I. Meeting adjourned by IBC Chair at 2:17 pm.

Next Meeting: September 3, 2026- Executive Board Room, Building 549 (WebEx access available).